

Functional Sequence Complexity (FSC) Measured in “Fits” (Functional bits).

Scirus Sci-Topics Page

David L. Abel, Director The Gene Emergence Project
Department of ProtoBioCybernetics & ProtoBioSemiotics
The Origin of Life Science Foundation, Inc.

Sequence complexity has three subsets: Random (RSC), Ordered (OSC), and Functional (FSC).¹ Functional Sequence Complexity is measured in “Fits.” Fits are “functional bits.”²⁻⁴

To understand Functional Sequence Complexity (FSC), one must first digest the essence of the other two subsets of sequence complexity. Random Sequence Complexity (RSC) lies at the opposite end of a bi-directional sequence complexity vector from Ordered Sequence Complexity (OSC).



Random Sequence Complexity (RSC) is defined by an inability to compress a sequence into a representation shorter than the sequence itself. The sequence lacks any redundant order or pattern that would allow compression. With RSC, no patterns exist in the sequence either from natural law constraints or from repeated use of programming modules.

Ordered Sequence Complexity (OSC) is typically produced by law-like cause-and-effect determinism. Such forced ordering produces boring redundancy and also precludes choice contingency needed for any form of programming. Combinatorial uncertainty, freedom of selection, and

potential information instantiation are all precluded in highly ordered strings. An example of a highly ordered string is a polyadenosine that adsorbs naturally onto montmorillonite clay.⁵⁻⁷ Algorithmic programming and control are made impossible when sequences are constrained by law.

Functional Sequence Complexity (FSC) is invariably associated with all forms of non-trivial formal utility. The algorithmic programming of FSC requires anticipation of the future. Purposeful choices for potential final function must be made. Mere aperiodicity of a sequence is not sufficient to define FSC. RSC is aperiodic; yet RSC produces no formal and final function.

Usually, FSC comes in the form of linear digital prescription using a symbol system. FSC requires the programming dimension of uncoerced choices for potential function at successive decision nodes in the string. RSC has mere bifurcation points, with nothing more than coin flips at each successive fork in the road to determine which fork to take. No expectation of improved trip efficiency exists when coin flips are used to determine which fork in the road to take.

Rats improve their exit time from mazes by memorizing wise purposeful choices at each successive decision node. Those choices must be made prior to the realization of any function. Utility (making it out of the maze) is only realized at the end of a long string of choices. The choices must be made IN PURSUIT OF eventual usefulness, not immediate gradification.

A succession of purposeful binary choices can be recorded as a string of symbols (e.g., 0's vs. 1's). That string of symbols represents FSC, the same as any computational program in computer science.

Thus, FSC arises only out of wise choices at true decision nodes, logic gates, and purposeful configurable switch-settings. The latter can only be set by formalism, not physicality, if sophisticated function is to be realized. In the generation of FSC, not only must each successive choice opportunity be free from physiodynamic determinism, it must be deliberately chosen en route to achieving eventual formal and final function.

Choice Determinism (CD), as opposed to Physiodynamic Determinism (PD), is quite real in producing any FSC string. The causation of FSC is formal (abstract, conceptual, and choice-based), not physical. The generation of FSC strings has never been observed to come into existence independent of agency. Zero empirical evidence exists of inanimate

physicality producing an integrated circuit, a genetic algorithm, a symbol system, language, or computational success.

No empirical evidence exists of either RSC or OSC ever having produced a single instance of sophisticated function or true organization. Algorithmic optimization requires purposeful choices to pursue eventual ideal function. Prescriptive Information (PI), circuit integration, and organization all invariably manifest FSC. Any attempt to deny the need and reality of purposeful choices precludes the production of any sophisticated function. Naturalistic philosophic presuppositions militate against acknowledging the obvious facts of reality. “Chance and Necessity” is a false dichotomy. Reality actually consists of three fundamental categories, not two: Chance, Necessity and Choice. By Choice, we do *not* mean mere Selection *FROM AMONG* [evolution]. The kind of Choice clearly observed everyday by everyone as a major component of reality includes Selection *FOR (IN PURSUIT OF)* not yet existent function.⁸⁻¹¹ Inanimate nature cannot exercise or generate such choice with intent. Only agency does. Mere mass/energy interactions have never been shown to produce the slightest hint of agency.

Nucleic acid genes, promoters and other regulators are examples of FSC, not OSC or RSC. The sequencing of nucleotides in single positive strands is physicodynamically indeterminate (free, unconstrained by natural law). Clearly this sequencing is not random either. Way too much sophisticated control is prescribed by all these sequences to attribute to noise. Meaningful/functional messages are also sent and received using FSC strings (“messenger molecules;” biopolymers).

From the perspective of amino-acid-sequence prescription alone, genomes are programmed using sophisticated noise-reducing block codes (Triplet codons prescribe each amino acid). The redundancy found in the codon table is misleading, however. Superimposed on triplet codon language is a second independent language involving hexamers. Intracellular languages are multi-layered, using the same symbols, but with different meanings and functions in conveyed simultaneously in each language. Coding is therefore multidimensional. The hexameric language prescribes Translational Pausing (TP).^{12,13} TP in turn determines correct folding of the polyamino chain at the back door of the ribosome to produce properly folded molecular-machine proteins.¹³ Both of these superimposed languages manifest FSC in the sequencing of nucleotides. Each locus in the string represents a quaternary choice from among four options.

Linear sequence complexity has received extensive study in many areas relating to Shannon's syntactic transmission theory.¹⁴⁻¹⁶ This theory pertains only to communication engineering. Linear complexity was further investigated by Kolmogorov, Solomonoff, and Chaitin.¹⁷⁻²¹ Compressibility became the measure of linear complexity in this school of thought. Hamming pursued the goal of noise-pollution reduction in Shannon's communication channel through redundancy coding.²² Communication engineering has improved by leaps and bounds.

Little progress has been made, however, in measuring and explaining *intuitive information*. This is especially true regarding the derivation through natural process of semantic instruction. The purely syntactic approaches to sequence complexity of Shannon, Kolmogorov, and Hamming have little or no relevance to "meaning." Shannon acknowledged this in the 3rd paragraph of his first famous paper right from the beginning of his research.¹⁵ Inadequacy is still very apparent in more recent attempts to define and measure functional complexity and information.²³⁻⁵⁹

Nucleic acid instructions reside in linear, digital, and resortable sequences.⁶⁰⁻⁶³ Replication is sufficiently mutable for evolution, yet conserved, competent, and repairable for heritability.⁶⁴

In life-origin science, attention usually focuses on a theorized pre-RNA World.⁶⁵⁻⁶⁸ RNA chemistry is extremely challenging in a prebiotic context. Ribonucleotides are difficult to make and activate (charge). Oligoribonucleotides are also extremely hard to form, especially without templating. The maximum length of such single strands in solution is usually only eight to ten monomers (mers). As a result, many investigators suspect that some chemical RNA analog must have existed^{69,70}. For our purposes here of discussing linear sequence complexity, let us assume adequate availability of all four ribonucleotides in a pre-RNA prebiotic molecular evolutionary environment. Any one of the four ribonucleotides could be polymerized next in solution onto a forming single-stranded polyribonucleotide. Let us also ignore in our model for the moment that the maximum achievable length of aqueous polyribonucleotides seems to be no more than eight to ten monomers (mers). Physicochemical dynamics do not determine *the particular sequencing* of these single-stranded, untemplated polymers of RNA. The selection of the initial "sense" sequence is largely free of natural law influences and constraints. *Sequencing is dynamically inert*⁷¹.

Initial sequencing of single-stranded RNA-like analogs is crucial to most life-origin models. Particular sequencing leads not only to a theorized self- or mutually-replicative primary structure, but to catalytic capability of

that same or very closely-related sequence. One of the biggest problems for the pre-RNA World model is finding sequences that can *simultaneously* self-replicate and catalyze needed metabolic functions. For even the simplest protometabolic function to arise, large numbers of such self-replicative and metabolically contributive oligoribonucleotides would have to arise at the same place at the same time.

Little empirical evidence exists to contradict the contention that untemplated *sequencing* is dynamically inert (physically arbitrary). We are accustomed to thinking in terms of base-pairing complementarity determining sequencing. It is only in researching the pre-RNA world that the problem of single-stranded metabolically functional sequencing of ribonucleotides (or their analogs) becomes acute. And of course highly-ordered templated sequencing of RNA strands on natural surfaces such as montmorillonite clay offers no explanation for biofunctional sequencing. The question is never answered, "From what source did the *template* derive its functional information?" In fact, no empirical evidence has been presented of a naturally-occurring inorganic or organic template that contains anything more than combinatorial uncertainty. No bridge has been established between combinatorial uncertainty and utility of any kind.

Increased frequencies of certain ribonucleotides, CG for example, are seen in *post-textual* reference sequences. This is like citing an increased frequency of "qu" in post-textual English language. The only reason "q" and "u" have a higher frequency of association in English is because of arbitrarily chosen rules, not laws, of the English language. Apart from linguistic rules, all twenty-six English letters are equally available for selection at any sequential decision node. But we are attempting to model a purely pre-textual, combinatorial, chemical-dynamic *theoretical* primordial soup. No evidence exists that such a soup ever existed. But assuming that all four ribonucleotides might have been equally available in such a soup, no such "qu" type *rule*-based linkages would have occurred chemically between ribonucleotides. They are freely resortable apart from templating and complementary binding. Weighted means of each base polymerization would not have deviated far from $p = 0.25$. Dimers seem to show some physical predilections. But longer stochastic ensembles seem randomly sequenced, with no prescriptive function.

When we introduce ribonucleotide availability realities into our soup model, we would not expect hardly any cytosine to be incorporated into the early genetic code. Cytosine is extremely difficult even for highly skilled chemists to generate.^{72,73} If an extreme paucity of cytosine existed in a primordial environment, uncertainty would have been greatly reduced.

Heavily weighted means of relative occurrence of the other three bases would have existed. The potential for recordation of prescriptive information would have been reduced by the resulting high probability and low uncertainty of base “selection.” Self-ordering would have prevailed over complexity.

All aspects of life manifest extraordinarily high quantities of prescriptive information. Any self-ordering (law-like behavior) or weighted-mean tendencies (e.g., reduced availability of certain bases) would have limited information instantiation and retention in the sequencing.

If non-templated chemistry predisposes higher frequencies of certain bases, how did so many highly-informational genes get coded? Any programming effort would have had to fight against a highly prejudicial self-ordering physycodynamic redundancy. There would have been little or no uncertainty (bits) at each locus. Information potential would have been severely constrained.

Functional Bits (Fits)

The evolution of amino acid sequence, and its effect on biofunction, can now be quantified in “fits” (functional bits).⁴

To understand how Functional Sequence Complexity can be measured, we must first understand “Functional Uncertainty (H_f):”

“Shannon's original formulation, when applied to biological sequences, does not express variations related to biological functionality such as metabolic utility. Shannon uncertainty, however, can be extended to measure *the joint variable* (X, F), where X represents the variability of data, and F functionality. This explicitly incorporates empirical knowledge of metabolic function into the measure that is usually important for evaluating sequence complexity. This measure of both the observed data and a conceptual variable of function jointly can be called *Functional Uncertainty* (H_f),⁷⁴ and is defined by the equation:

$$H(X_f(t)) = - \sum P(X_f(t)) \log P(X_f(t)) \quad (1)$$

where t = a certain time and X_f denotes the conditional variable of the given sequence data (X) on the described biological function f which is an outcome of the variable (F).⁷⁴

In this approach, f might represent the known 3-D structure of a protein family. The entire set of aligned sequences that satisfies that protein's function, therefore, would constitute the outcomes of X_f . The advantage of using $H(X_f(t))$ is that evolutionary changes through time in the functionality of sequences can be measured.

Functional uncertainty as a measure of FSC

The measure of Functional Sequence Complexity, denoted as ζ , is defined as the change in functional uncertainty from the ground state $H(X_g(t_i))$ to the functional state $H(X_f(t_i))$, or

$$\zeta = \Delta H (X_g(t_i), X_f(t_j)) . \quad (2)$$

The resulting unit of measure is defined on the joint data and functionality variable, which we call *Fits* (or *Functional bits*). The unit Fit thus defined is related to the intuitive concept of *functional* information, including genetic instruction and, thus, provides an important distinction between functional information and Shannon information.^{75,76}

The limitation of Functional Sequence Complexity (FSC) measurements is that they are nonspecific averages. In addition, the change in negative Shannon Uncertainty is only obtained by the extrinsic injection of true positive information into the equation. Our probabilistic combinatorial uncertainty is educated only by the empirical data providing the relative certainty of which particular sequences will work. We sneak in through the back door, in other words, the real semantic, functional information rather than the equation generating it. The empirical data is only obtained after the fact, and in very general statistical terms.

The reason FSC does not qualify as Prescriptive Information (PI)⁷⁷ is that it cannot specifically enumerate which particular sequences will work. The latter is the real essence of intuitive, functional, prescriptive information (PI).

References

1. Abel DL, Trevors JT. Three subsets of sequence complexity and their relevance to biopolymeric information. *Theoretical Biology and Medical Modeling*. 2005;2:29-45.
2. Durston KK. Methods: Measuring the 'Functional Sequence Complexity' of proteins: *Methods*. 2007; <http://www.uoguelph.ca/~kdurston/Methods.rtf>.

3. Durston KK. Python program to analyze the two-dimensional array of aligned sequences for a protein family. 2007; <http://www.uoguelph.ca/~kdurston/>. Accessed November 13, 2007.
4. Durston KK, Chiu DK, Abel DL, Trevors JT. Measuring the functional sequence complexity of proteins. *Theoretical biology & medical modelling*. 2007;4:Free on-line access at <http://www.tbiomed.com/content/4/1/47>.
5. Ferris JP, Huang CH, Hagan WJ, Jr. Montmorillonite: a multifunctional mineral catalyst for the prebiological formation of phosphate esters. *Orig Life Evol Biosph*. 1988;18(1-2):121-133.
6. Ferris JP, Ertem G. Oligomerization of ribonucleotides on montmorillonite: reaction of the 5'-phosphorimidazolidine of adenosine. *Science*. 1992;257(5075):1387-1389.
7. Bujdak J, Eder A, Yongyai Y, Faybikova K, Rode BM. Investigation on the mechanism of peptide chain prolongation on montmorillonite. *J Inorg Biochem*. Jan 1996;61(1):69-78.
8. Abel DL. The three fundamental categories of reality. In: Abel DL, ed. *The First Gene: The Birth of Programming, Messaging and Formal Control*. New York, N.Y.: LongView Press-Academic: Biol. Res. Div.; 2011:19-54 Also available from <http://lifeorigin.academia.edu/DrDavidLAbel>.
9. Abel DL. The Cybernetic Cut and Configurable Switch (CS) Bridge. In: Abel DL, ed. *The First Gene: The Birth of Programming, Messaging and Formal Control*. New York, N.Y.: LongView Press--Academic, Biol. Res. Div.; 2011:55-74 Also available from <http://lifeorigin.academia.edu/DrDavidLAbel>.
10. Abel DL. What utility does order, pattern or complexity prescribe? In: Abel DL, ed. *The First Gene: The Birth of Programming, Messaging and Formal Control*. New York, N.Y.: LongView Press--Academic, Biol. Res. Div.; 2011:75-116 Also available from <http://lifeorigin.academia.edu/DrDavidLAbel>.
11. Abel DL. *Primordial Prescription: The Most Plaguing Problem of Life Origin Science*. New York, N.Y.: LongView Press Academic; 2015.
12. Li G-W, Oh E, Weissman JS. The anti-Shine-Dalgarno sequence drives translational pausing and codon choice in bacteria. *Nature*. 2012;484(26 APRIL):538
13. D'onofrio DJ, Abel DL. Redundancy of the genetic code enables translational pausing. *Frontiers in Genetics*. 2014-May-20 2014;5:140 Open access at <http://www.ncbi.nlm.nih.gov/pmc/articles/PMC4033003/>.
14. Shannon CE, Weaver W. *The Mathematical Theory of Communication*. Urbana, IL: University of Illinois Press; 1949.
15. Shannon C. Part I and II: A mathematical theory of communication. *The Bell System Technical Journal*. 1948;XXVII(3 July):379-423.
16. Shannon C. Part III: A mathematical theory of communication. *The Bell System Technical Journal*. 1948;XXVII(3 July):623-656.
17. Kolmogorov AN. Three approaches to the quantitative definition of the concept "quantity of information". *Problems Inform. Transmission*. 1965;1:1-7.
18. Li M, Vitanyi P. *An Introduction to Kolmogorov Complexity and Its Applications*. 2 ed. New York: Springer-Verlag; 1997.

19. Chaitin GJ. Algorithmic information theory. *IBM Journal of Research and Development*. 1977;21:350-359.
20. Chaitin GJ. *Computational complexity and Godel's incompleteness theorem and To a mathematical definition of life*. [Rio de Janeiro,: Centro Tâecnico Cientâifico, Pontifâicia Universidade Catâolica do Rio de Janeiro; 1970.
21. Chaitin GJ. On the length of programs for computing finite binary sequences. *J. ACM*. 1966;13:547.
22. Hamming RW. *Numerical methods for scientists and engineers*. 2nd ed. New York: Dover; 1986.
23. MacKay DM. *Information, Mechanism and Meaning*. Cambridge, MA: M.I.T. Press; 1969.
24. Badii R, Politi A. *Complexity : hierarchical structures and scaling in physics*. Cambridge ; New York: Cambridge University Press; 1997.
25. Adami C, Cerf NJ. Physical complexity of symbolic sequences. *Physica D*. 2000;137:62-69.
26. Adami C, Ofria C, Collier TC. Evolution of biological complexity. *Proc Natl Acad Sci U S A*. 2000;97(9):4463-4468.
27. Adami C. What is complexity? *Bioessays*. 2002;24(12):1085-1094.
28. Allison L, Stern L, Edgoose T, Dix TI. Sequence complexity for biological sequence analysis. *Comput Chem*. 2000;24(1):43-55.
29. Bennett DH. Logical depth and physical complexity. In: Herken R, ed. *The Universal Turing Machine: a Half-Century Survey*. Oxford: Oxford University Pres; 1988.
30. Bennett CH. How to define complexity in physics, and why. In: Zurek WH, ed. *Complexity, Entropy and the Physics of Information, SFI studies in the Sciences of Complexity*. Vol 81990:137-148.
31. Dewey TG. Algorithmic complexity of a protein. *Physical Review. E. Statistical Physics, Plasmas, Fluids, and Related Interdisciplinary Topics*. 1996;54(1):R39-R41.
32. Gell-Mann M. What is complexity? *Complexity*. 1995;1(1):16-19.
33. Gell-Mann M, Lloyd S. Information measures, effective complexity, and total information. *Complexity*. 1996;2(1):44-52.
34. Kauffman S. *At Home in the Universe: The Search for the Laws of Self-Organization and Complexity*. New York: Oxford University Press; 1995.
35. Konopka AK, Owens J. Complexity charts can be used to map functional domains in DNA. *Genet Anal Tech Appl*. 1990;7(2):35-38.
36. Konopka AK. Sequences and Codes: Fundamentals of Biomolecular Cryptology. In: Smith D, ed. *Biocomputing: Informatics and Genome Projects*. San Diego: Academic Press; 1994:119-174.
37. Konopka AK. Sequence complexity and composition. In: Cooper DN, ed. *Nature Encyclopedia of the Human Genome. Vol. 5*. London: Nature Publishing Group Reference; 2003:217-224.
38. Lempel A, Ziv J. On the complexity of finite sequences. *IEEE Trans Inform. Theory*. 1976;22:75.
39. Lenski RE, Ofria C, Collier TC, Adami C. Genome complexity, robustness and genetic interactions in digital organisms. *Nature*. 1999;400(6745):661-664.

40. Nicolaou KC. Creating complexity – the beauty and logic of synthesis. *Chem. Commun.* 2003;6:661 - 664.
41. Ricard J. What do we mean by biological complexity? *C R Biol.* 2003;Feb;326(2):133-140.
42. Weng G, Bhalla US, Iyengar R. Complexity in biological signaling systems. *Science.* 1999;284(5411):92-96.
43. Zurek WH. Thermodynamic cost of computation, algorithmic complexity, and the information metric. *Nature.* 1989;341:119-124.
44. Pincus SM. Irregularity and asynchrony in biologic network signals. *Methods Enzymol.* 2000;321:149-182.
45. Conrad M. Unity of measurement and motion. *Biosystems.* 2001;60:23-38.
46. Rosen R. *Essays On Life Itself.* New York: Columbia University Press; 2000.
47. Rosen R. *Life Itself: A Comprehensive Inquiry Into the Nature, Origin and Fabrication of Life.* New York: Columbia University Press; 1991.
48. Mikulecky DC. The emergence of complexity: science coming of age or science growing old? *Computers Chem.* 2001;25:341-348.
49. Mikulecky DC. Network thermodynamics and complexity: a transition to relational systems theory. *Comput Chem.* Jul 2001;25(4):369-391.
50. Lewontin R. *The Triple Helix: Gene, Organism and Environment.* Cambridge, MA: Harvard University Press; 2000.
51. Lewontin RC. Evolution as engineering. In: Collado-Vides J, Smith T, Magasanik B, eds. *Integrative Approaches to Molecular Biology.* Cambridge, MA: MIT Press; 1996.
52. Gordon R. Evolution Escapes Rugged Fitness Landscapes by Gene or Genome Doubling: the Blessing of Higher Dimensionality. *Computers Chem.* 1994;18:325 - 332.
53. Konopka AK. Grand metaphors of biology in the genome era. *Computers & Chemistry.* 2002;26:397-401.
54. Lewontin R, Levins R. Let the numbers speak. *International journal of health services.* 2000;30(4):873-877.
55. Bar-Hillel Y, Carnap R. Semantic Information. *British Journal for the Philosophy of Science.* 1953;4:147-157.
56. Mayr E. *This Is Biology: The Science of the Living World.* Cambridge, MA: Harvard University Press; 1997.
57. Bell GI. Evolution of simple sequence repeats. *Computers Chem.* 1996;20:41 - 48.
58. Reidys C, Stadler PF. Bio-molecular shapes and algebraic structures. *Computers Chem.* 1996;20:85 - 94.
59. Conrad M. Origin of life and the underlying physics of the universe. *Biosystems.* 1997;42(2-3):177-190.
60. Yockey HP. An application of information theory to the Central Dogma and the Sequence Hypothesis. *J Theor Biol.* 1974;46(2):369-406.
61. Yockey HP. *Information Theory and Molecular Biology.* Cambridge: Cambridge University Press; 1992.

62. Yockey HP. Informatics, Information Theory, and the Origin of Life. Paper presented at: 4th International Conference on Computational Biology and Genome Informatics 2002; Duke University; Research Triangle Park.
63. Yockey HP. Origin of life on earth and Shannon's theory of communication. *Comput Chem.* 2000;24(1):105-123.
64. Maeshiro T, Kimura M. The role of robustness and changeability on the origin and evolution of genetic codes. *PNAS.* 1998;95(9):5088-5093.
65. Lazcano A, Miller SL. The origin and early evolution of life: prebiotic chemistry, the pre-RNA world, and time. *Cell.* 1996;85(6):793-798.
66. Kolb VM, Dworkin JP, Miller SL. Alternative bases in the RNA world: the prebiotic synthesis of urazole and its ribosides. *J Mol Evol.* 1994;38:549-557.
67. Dworkin JP, Lazcano A, Miller SL. The roads to and from the RNA world. *J Theor Biol.* May 7 2003;222(1):127-134.
68. Joyce GF, Orgel LE. Prospects for understanding the origin of the RNA World. In: Gesteland RF, Cech TR, Atkins JF, eds. *The RNA World*. Second ed. Cold Spring Harbor, NY: Cold Spring Harbor Laboratory Press; 1999:49-78.
69. Monnard PA, Deamer DW. Preparation of vesicles from nonphospholipid amphiphiles. *Methods Enzymol.* 2003;372:133-151.
70. Kozlov IA, Orgel LE. Oligomerization of deoxyguanosine 5'-phosphoro-2-methylimidazolide on a polycytidylate template. *Orig Life Evol Biosph.* 1999;29(6):593-595.
71. Rocha LM. Evolution with material symbol systems. *Biosystems.* 2001;60:95-121.
72. Shapiro R. Comments on 'Concentration by Evaporation and the Prebiotic Synthesis of Cytosine'. *Origins Life Evol Biosph.* 2002;32(3):275-278.
73. Shapiro R. Prebiotic cytosine synthesis: a critical analysis and implications for the origin of life. *Proc Natl Acad Sci U S A.* 1999;96(8):4396-4401.
74. Durston KK, Chiu DKY. A functional entropy model for biological sequences. *Dynamics of Continuous, Discrete & Impulsive Systems, Series B.* 2005.
75. Szostak JW. Functional information: Molecular messages. *Nature.* Jun 12 2003;423(6941):689.
76. Barbieri M. *The Organic Codes: An Introduction to Semantic Biology*. Cambridge: Cambridge University Press; 2003.
77. Abel DL. The biosemiosis of prescriptive information. *Semiotica.* 2009;2009(174):1-19 Also available from <http://lifeorigin.academia.edu/DrDavidLabel>